

ACKNOWLEDGEMENTS

This study was supported by the Tennessee Wildlife Resources Agency from Federal Aid in Wildlife Restoration Project W-46-3 Tennessee, Department of Forestry, Agricultural Experiment Station, McIntire Stennis Project No. 11, and the Great Smoky Mountains Natural History Association.

LITERATURE CITED

- Belden, R. C. and M. R. Pelton. 1975. European wild hog rooting in the mountains of East Tennessee. Proc. 29th Ann. SE Game and Fish Comm. Conf. (In Press.)
- Cain, S. A. 1931. Ecological studies of the vegetation of the Great Smoky Mountains of North Carolina and Tennessee. I. Soil reaction and plant distribution. Bot. Gaz. 91:22-41.
- Dunne, H. W. ed. 1970. Diseases of swine (3rd ed.). Iowa State Univ. Press, Ames. 1144 pp.
- Fox, J. R. 1972. An evaluation of control techniques for the European wild hog in the Great Smoky Mountains National Park of Tennessee. M.S. Thesis. Univ. of Tennessee, Knoxville, 76 pp.
- Jones, P. 1959. The European wild boar in North Carolina. Game Div., North Carolina Wildl. Resour. Comm., Raleigh. 29 pp.
- Mount, L. E. 1968. Adaptation of swine. Pages 277-291 in E. S. E. Hafez, ed., Adaptation of domestic animals. Lea and Febiger, Philadelphia. 415 pp.
- Shanks, R. E. 1954a. Reference list of native plants in the Great Smoky Mountains. Botany Dept., University of Tennessee. Mimeo. 14 pp.
- Shanks, R. E. 1954b. Climates of the Great Smoky Mountains. Ecology 35(3):354-361.
- Stegeman, L. C. 1938. The European wild boar in the Cherokee National Forest, Tennessee. J. Mammal. 19(3):279-290.
- U.S. Forest Service. 1970. Tellico District multiple use plan. Chap. 100:150-151.
- Whittaker, R. H. 1956. Vegetation of the Great Smoky Mountains. Ecol. Monogr. 26:1-80.

JOURNAL OF THE TENNESSEE ACADEMY OF SCIENCE
VOLUME 51, NUMBER 3, JULY, 1976

SYNTHESIS AND STUDIES OF BACTERIOSTATIC ACTIVITY OF SULFILIMINES

JITENDRA J. SHAH
Woodson-Tenent Laboratories, Inc.
Memphis, Tennessee 38101

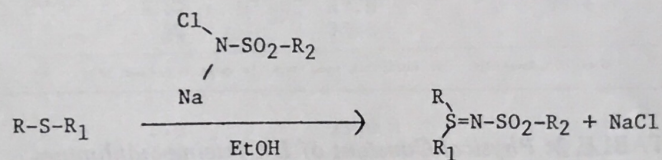
ABSTRACT

A new series of Amino Acids-Sulfilimine have been prepared. They were tested against *Escherichia coli* and *Sarcina lutea* in presence of an antibiotic medium. Also studied was the toxicity in mice.

INTRODUCTION

The amino acids, L-Methionine and L-Cysteine recently have been found to react with Chloramine-B, Chloramine-T and N-chloro-N-sodio-*p*-ethyl benzene-sulfonamide to obtain the corresponding sulfilimines. Inhibition of the growth of *Escherichia coli* and *Sarcina lutea* by these amino acid derivatives of sulfilimines have been described by others. (Dakin, Cohen, *et al*, derivatives may function as competitive inhibitors in the metabolism of the corresponding amino acids. The acute oral toxicity of some amino acid derivatives have been determined in male albino mice.

Several N-*p*-tolyl sulfonyl derivatives of sulfilimines have been described by others. (Dakin, Cohen, *et al*, 1916; Inglis, 1918; and Nicolet and Willard, 1921). Recently there has been prepared a new Chloramine (N-chloro-N-sodio-*p*-ethyl benzene sulfonamide) (Shah and Claypool, 1972) and a new series of sulfilimines called N-*p*-ethyl benzene sulfonyl sulfilimines. The sulfilimines of L-Methionine are given in Table 1, were prepared by treating the corresponding sulfide in the N-sulfonyl ha'omide reaction.



Compound # 1, R= -CH₃, R₁= -CH₂-CH₂- $\begin{array}{c} \text{H} \\ | \\ \text{C-COOH} \\ | \\ \text{NH}_2 \end{array}$, R₂= -C₆H₅

2, " " R₂= -C₇H₇

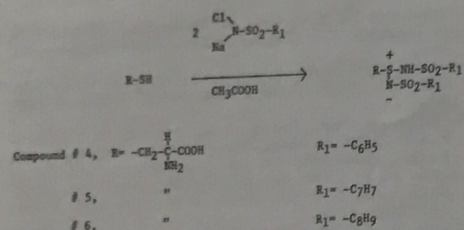
3, " " R₂= -C₈H₉

The sulfilimines of L-Cysteine are given in Table 2, were prepared by the action on N-sulfonyl halomide on solutions of mercaptans in glacial acetic acid.

These amino acid derivatives were tested against *Escherichia coli* and *Sarcina lutea* in the presence of antibiotic medium (I) and (IV), pH 6 phosphate buffer, with 16-20 hr incubates at 30-31°C. This method was first described by Abraham *et al*, (1941) for the assay of Penicillin. It was later modified by Foster and Woodruff (1943) and by Schmidt and Moyer. (1944) The acute oral LD₅₀ for compounds (2 & 5) are greater than 5.0 gm/kg of body weight.¹

In summary, analytical data are not adequate proof for

¹ Hill-Top Research, Inc., Miami, Ohio 45147.



the sulfilimines prepared but the spectral data verify the preparation of the new sulfilimines in this work. Such verification has been successfully utilized by K. Tsujihara (1969) in his work with sulfilimines. The diameter of the zones are directly dependent upon the concentration of the antimicrobial agents. Figures 1 & 2 show that the inhibition of *Sarcina lutea* and *Escherichia coli* on L-Methionine & L-Cysteine-sulfilimines. From these

TABLE 1: Physical Constants of L-Methionine-Sulfilimines

Compound #	R	mp, °C	lit. ref.	Infrared, cm ⁻¹	UV-visiblelet NaOH x max	Analysis
(1)	-R	138-139	25	1710, 1077 1170 925	259, 262 267, 272	C ₁₁ H ₁₆ N ₂ O ₄
(2)	-CH ₂	134-135	29	1295, 1030 1167 928	256, 262 268, 273	C ₁₀ H ₁₄ N ₂ O ₄
(3)	-C ₂ H ₅	98-99	18	1203, 1032 1164 930	254, 261 267, 272	C ₁₂ H ₁₈ N ₂ O ₄

²⁵o: Calcd, 23.5; Found 22.9. ²⁹i: Calcd, 22.3; Found, 21.8. ¹⁸o: Calcd, 20.8; Found, 20.3.

TABLE 2: Physical Constant of L-Cysteine Sulfilimines

Compound #	R ₁ & R ₂	mp, °C	lit. ref.	Infrared, cm ⁻¹	UV-visiblelet (CHCl ₃) x max	Analysis
(4)	-R	136-136.5	70	1220, 1060 1164 932	257, 261 269, 274	C ₁₂ H ₁₈ N ₂ O ₄
(5)	-CH ₂	138	65	1218, 1038 1162 930	256, 263 268, 273	C ₁₁ H ₁₄ N ₂ O ₄
(6)	-C ₂ H ₅	107-108	45	1211, 1030 1160 931	255, 262 268, 273	C ₁₃ H ₁₈ N ₂ O ₄

⁷⁰i: Calcd, 23.5; Found, 22.9.

figures the critical concentration of each compound can be determined. (Kavanagh, 1963) The Penicillin G Potassium readily inhibits *Sarcina lutea* at a very low concentration (0.025 μg/ml), but at high concentration (0.1 μg/ml) inhibits *Escherichia coli*. These amino acid derivatives inhibit *Sarcina lutea* at a low concentration (30 μg/ml), but at a high concentration

(50 μg/ml) inhibits *Escherichia coli*. Amino acids themselves do not inhibit either *Sarcina lutea* or *Escherichia coli*.

These amino acid derivatives are (1200:1) more than a thousand times less active than the Penicillin G Potassium in *Sarcina lutea*, and (500:1) 500 times less active than Penicillin G Potassium in *Escherichia coli*. These results suggest that these amino acid derivatives have some antimicrobial and toxic properties.

MATERIALS AND METHODS

Sulfilimines of analytical purity as listed in Table 1 & 2 were prepared from appropriate Chloramine and Sulfides or Mercaptans either by known methods (cf Tables) or by the following general procedure.

(a) *From Sulfides*: (McCall, Tarbell and Havill, 1951): The mixed solutions of 0.02 mol of sulfide (in 50 ml ethyl alcohol) and of 0.03 mol of chloramine (in 50 ml of 50% ethyl alcohol-water solution) were heated in a water bath for 30 minutes. It was then covered and allowed to stand overnight at room temperature. When product formed on standing the product was filtered, washed thoroughly with water, dried and recrystallized from EtOH or MeOH (once or twice).

(1) *Methionine-benzene sulfonylimine*:— The recrystallized product was obtained by the general procedure outlined above; yield 25%, mp 138-139°, ir (CHCl₃): (Asymm SO₂): 1210, 1027 cm⁻¹, (Symm SO₂): 1170 cm⁻¹, (S=N) 925 cm⁻¹. Anal. calcd for C₁₁H₁₆N₂O₄: C, 48.60; H, 5.88; N, 10.29; S, 23.5. Found, C, 48.60; H, 5.73; N, 9.98; S, 22.9.

(2) *Methionine-p-toluene-sulfonylimine*:— The recrystallized product was prepared by the general procedure; yield 28%, mp 134-135°, ir (CHCl₃): (Asymm SO₂): 1203, 1030 cm⁻¹, (Symm SO₂): 1167 cm⁻¹, (S=N): 928 cm⁻¹. Anal. calcd for C₁₂H₁₈N₂O₄: C, 50.35; H, 6.30; N, 9.80; S, 22.3. Found: C, 50.10; H, 5.98; N, 9.98; S, 21.8.

(3) *Methionine-p-ethyl benzene sulfonylimine*:— The recrystallized product was prepared by the general procedure; yield 18%, mp 98-99°, ir (CHCl₃): (Asymm SO₂): 1230, 1032 cm⁻¹, (Symm SO₂): 1166 cm⁻¹, (S=N): 930 cm⁻¹. Anal. calcd for C₁₃H₂₀N₂O₄: C, 52.00; H, 6.67; N, 9.30; S, 20.8. Found: C, 52.40; H, 6.50; N, 8.98; S, 20.3.

From Mercaptans—(Clark, Kenyon and Phillips, 1930): The mixed solutions of 1.10 mol of mercaptan (in 200 ml glacial acetic acid) and of 1.1452 mol of chloramine (in 100 ml glacial acetic acid) were heated in water bath for 20 min., the mixture was poured into water. The solid product which separated was extracted with benzene to remove disulfide, and then digested with hot dilute sulfuric acid. A heavy viscous oil remained which crystallized on cooling and recrystallized from EtOH or MeOH (twice).

(4) *Cysteine-benzene sulfonylimido sulfine-benzene-sulfonylimine*:— The recrystallized product was prepared by the general procedure; yield 70%, mp 136-136.5°, ir (CHCl₃): (Asymm SO₂): 1200, 1040 cm⁻¹, (Symm SO₂): 1168 cm⁻¹, (S=N): 932 cm⁻¹. Anal. calcd for C₁₂H₁₇N₂O₄: C, 44.30; H, 5.10; N, 8.60; S, 22.5. Found: 44.70; H, 4.86; N, 8.07; S, 21.4.

(5) *Cysteine-p-toluene sulfonylimido sulfine-p-toluene sulfonylimines*:— The recrystallized product was prepared by the general procedure; yield 65%, mp 138°, ir (CHCl₃): (Asymm SO₂): 1218, 1038 cm⁻¹, (Symm SO₂): 1167 cm⁻¹, (S=N): 930 cm⁻¹. Mol. Formula: C₁₁H₁₄N₂O₄.

(6) *Cysteine-p-ethyl benzene sulfonylimido sulfine-p-ethyl benzene sulfonylimine*:— The recrystallized product was prepared by the general procedure; yield 45%, mp 107-108°, ir (CHCl₃): (Asymm SO₂): 1211, 1030 cm⁻¹, (Symm SO₂): 1160 cm⁻¹, (S=N): 921 cm⁻¹. Mol. Formula: C₁₃H₁₈N₂O₄.

MICROBIOLOGICAL TEST²

Accurately weigh ca. 10 mg of each amino acid derivative in separate 100 ml volumetric flask and dissolve in 10 ml HCl and enough pH 6 phosphate buffer to give an exact concentration of 100 μg/ml (solution a). Dilute approximate aliquots of solution (a), with enough pH 6 buffer to obtain concentrations of 70-, 60-, 50-, 40-, 30 μg/ml. Add 10 ml melted Bacto-antibiotic-medium I to sterile petri dishes, distribute evenly and let harden on perfectly level surface. For actual assay approximate amounts of organism-suspensions are added to (1 ml) Bacto-antibiotic-medium-IV previously melted and cooled to 48°C. Mix thoroughly and add 4.0 ml to each plate containing base layer of Bacto antibiotic-medium-I. Distribute media evenly by tilting plates from side to side with circular motion and let harden.

Place three cylinders on each plate at ca 60° intervals on a 2.8 cm radius. Fill all 3 cylinders with the test solution. Incubate plates overnight at 30-31°C and measure the diameters of zones of inhibition by means of mm ruler, calipers or calibrated projection device. Three plates were used for each assay solution and three plates for the standard solution. Determine the corrected value of the sample and standard. Plotting values of x² (square of the zone size) against ln m⁰ (logarithm of the concentration in the reservoir) gives a straight line intercepting the concentration axis at ln m_c (critical concentration).

TOXICITY TESTS

Each sample was administered to male albino mice (Laboratory Supply-Company), weight range 18 to 27 gms, at a dosage level of 5.0 gm/kg of body weight. The samples were administered as 50% weight/volume suspensions in corn oil (Mazola).

Food was withheld from the mice for approximately 18 hrs. prior to dosage. Following dosage, food consisting of commercial pellets and water were available *ad libitum*. The mice were housed in conventional box-type mouse cages in groups of ten. All animals were observed closely for gross signs of systemic toxicity and mortality on the day of dosage, and at least once daily thereafter for a total of 14 days. Gross necropsies were performed on the animals that died. At the end of the 14-day observation period the surviving mice were weighed, sacrificed by cervical dislocation and gross necropsies were performed.

With compound # 2, no mortalities occurred. Therefore the acute oral LD₅₀ for male albino mice is greater than 5.0 gm/kg of body weight. On the day of dosage the mice appeared depressed and showed depressed righting and placement reflexes, ataxia and a rapid shallow respiration. On the following day and for the remainder of the 14-day observation period the mice exhibited normal behavior and appearance. The mice showed an average weight gain of seven gms which is within normal limits for mice of the age, sex and strain used in this study. Gross necropsies performed at termination showed no gross pathology.

In compound #5, three mice died during the study. Two deaths occurred within two hrs. of dosage. The third death occurred on the third-post dosage day. Therefore, the acute oral LD₅₀ for male albino mice is greater than 5.0 gm/kg of body weight. On the day of dosage the mice appeared depressed and exhibited depressed righting and placement reflexes, ataxia and rapid shallow respiration. These signs persisted without substantial change through the second post-dosage day. On the third-post-dosage day and for the remainder of the 14-day observation period the surviving mice appeared grossly normal. The mice showed an average body weight gain of eight gms

TABLE 3: Microbiological Data of Sulfilimines

Compound #	Concentration m ₀ (μg/ml)	<i>Sarcina lutea</i> Diameter of zones x (mm)	Critical-Concentration m ¹ (μg/ml)	<i>Escherichia coli</i>		Critical-Concentration m ¹ (μg/ml)
				Concentration m ₀ (μg/ml)	Diameter of Zones x (mm)	
(1)	50	17.1	21.5	70	21.0	39.4
	40	14.6		60	17.6	
	30	12.1		50	13.8	
(2)	50	18.1	22.5	70	19.0	32.0
	40	15.2		60	16.6	
	30	12.2		50	14.2	
(3)	50	17.4	20.9	70	21.6	31.8
	40	14.9		60	18.0	
	30	12.3		50	14.8	
(4)	50	17.8	24.0	70	21.0	34.8
	40	14.75		60	18.0	
	30	11.8		50	14.8	
(5)	50	17.8	18.5	70	20.1	38.0
	40	15.2		60	17.0	
	30	12.8		50	13.8	
(6)	50	17.8	18.0	70	21.4	38.0
	40	14.6		60	17.8	
	30	12.5		50	14.3	

² (Horwitz, 1970)

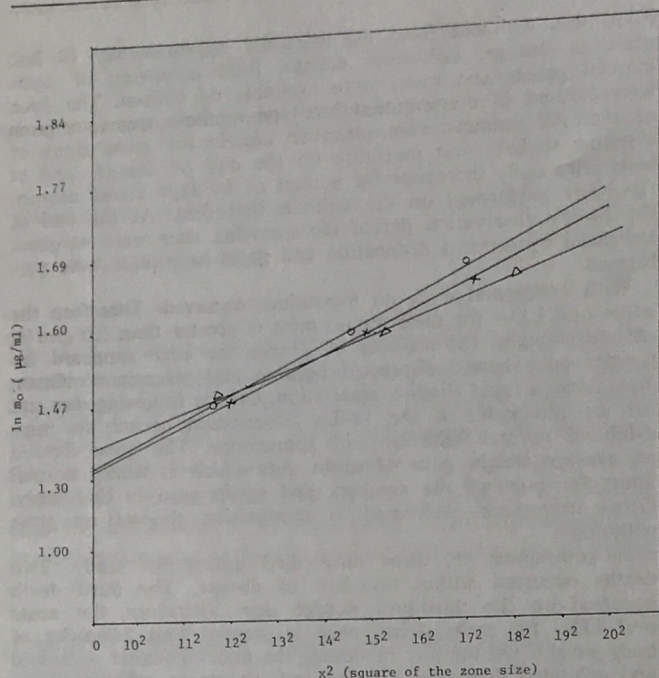


Fig. 1: Inhibition of *Sarcina lutea* on *L*-Methionine-sulfilimines.

- — ○ Methionine-benzene-sulfonylimine
- △ — △ Methionine-p-toluene-sulfonylimine
- x — x Methionine-p-ethyl benzene sulfonylimine

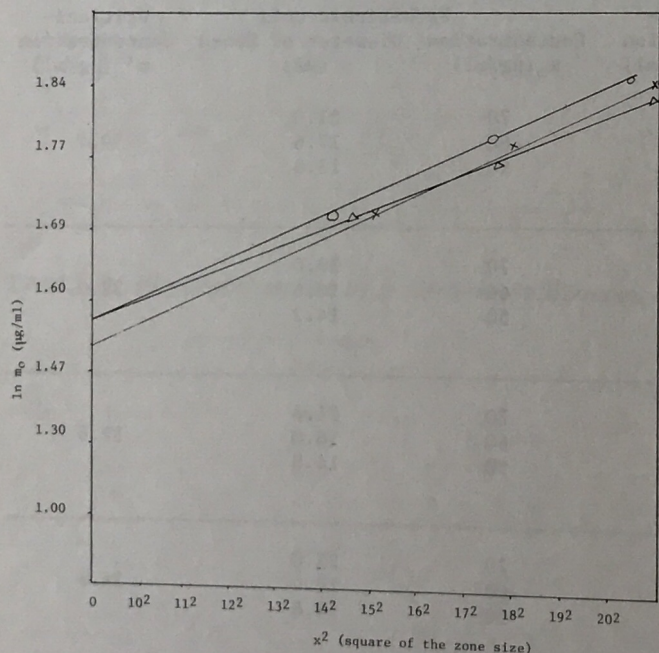


Fig. 2: Inhibition of *Escherichia coli* on *L*-Cysteine-sulfilimines.

- x — x Cysteine-benzene sulfonimido sulfine-benzene-sulfonylimine
- — ○ Cysteine-p-toluene sulfonimido sulfine-p-toluene sulfonylimine
- △ — △ Cysteine-p-ethyl benzene sulfonimido sulfine-p-ethyl benzene sulfonylimine

which is within normal limits for mice of the age, sex and strain used in this study. Gross necropsies performed on the surviving mice at termination showed no gross pathology.

ACKNOWLEDGEMENT

The author wishes to thank the Woodson-Tenent Laboratories, Inc. for their cooperation and research facilities.

LITERATURE CITED

Abraham, E. P. 1941. Growth of Penicillin Producing Mould, *Lancet* 2:177.

Clarke, S. G., Kenyon, J. and Phillips, H. 1927. Investigation on The Dependence of Rotatory Power on Chemical Constitution. *J. Chem. Soc.* 188.

Dakin, H. D., Cohen, J. R. and others. 1916. Studies in Antiseptics II "Chloramine," its Preparation, Properties and Use. *J. Brit. Med.* 160.

Foster, J. W. and Woodruff, H. B. 1943. Microbiological Aspects of Penicillin. *J. Bact.* 46:187.

Horwitz, W. 1970. Determination of Procaine Penicillin in Feeds. "Official Methods of Analysis of A.O.A.C." 11th Ed. Washington, D.C. 761 p.

Inglis, J. K. H. 1918. The Manufacture of Chloramine-T. *J. Soc. Chem. Ind.* 37:288T.

Kavanagh, F. 1963. The Theory of Antibiotic Inhibition Zones. "Analytical Microbiology." Academic Press 13 p.

McCall, M. A., Tarbell, D. S. and Havill, Mary, Ann. 1951. The Hydrogenolysis of Sulfilimines and its Application to The Purification of Sulfides. *J. Am Chem. Soc.* 4477.

Nicolet, B. H. and Willard, D. J. 1921. Preparation of Sulfilimines, *Science* 53:217.

Schmidt, W. H. and Moyer, A. J. 1944. Penicillin (Methods of Assay). *J. Bact.* 47:199.

Shah, J. J. 1972. Synthesis and Their Effect Upon The Growth of *Escherichia Coli* and *Sarcina Lutea* on Iminosulfuranes. 164th National Meeting of The American Chemical Society, New York. #BIOL 232.

Shah, J. J. and Claypool, D. P. 1972. Synthesis and Spectral Studies of Iminosulfuranes and Their P. As and Sb Analogous Compounds. 28th South West Regional Meeting of The American Chemical Society, Baton Rouge, Louisiana #175.

Shah, J. J. 1971. (M.S. Thesis). The Synthesis and Reactions of Iminosulfuranes (Sulfilimines) and Their Analogous Compounds. Memphis State University.

Tsujihara, K.; Furukawa, N.; Oae, K. and Oae, S. 1969. Sulfilimines II. IR, UV and NMR Spectroscopic Studies. *Bulletin of The Chemical Society of Japan.* 42:2631.